

Validation of the folate food frequency questionnaire in vegetarians

IRENA COLIĆ BARIĆ¹, ZVONIMIR ŠATALIĆ¹, ŽELJKO PEDIŠIĆ²,
VESNA ŽIZIĆ³ & IRENA LINARIĆ³

¹Faculty of Food Technology and Biotechnology, University of Zagreb, Zagreb, Croatia, ²Faculty of Kinesiology, University of Zagreb, Zagreb, Croatia, and ³Children's Hospital Zagreb, Zagreb, Croatia

Abstract

Adequate folate status has an important role in the prevention of chronic and developmental diseases and is considered a potential public health issue. Therefore, valid tools for measuring the vitamin intake are needed. In our previous study a folate food frequency questionnaire¹ (FFQ) designed to measure dietary folate equivalents was developed and validated among adult women against serum and erythrocyte (red blood cell) folate and plasma homocysteine. The aim of the present study was to validate the FFQ in vegetarians ($n=75$). The Pearson correlation for folate intake and biomarkers was 0.41, 0.36 and -0.15 for serum and red blood cell folate and plasma homocysteine, respectively. The quadratic weighted kappa value for biomarkers was above 0.2 and the gross misclassification of subjects into quartiles was less than 10%. The FFQ is a valid tool for measuring dietary folate equivalent intake in Croatian vegetarians.

Keywords: Food frequency questionnaire, validation, folate, vegetarians

Introduction

Folate is the generic term for a water-soluble B complex vitamin that functions in single-carbon transfer reactions and exists in many chemical forms (Wagner 1996). The nutritional status of the vitamin is considered a potential public health issue requiring further study (Life Sciences Research Office 1989).

Adequate folate intake has an important role in the primary prevention of several diseases (Bailey and Gregory 2006), and adequate folate intake is especially important during the periconceptional period as a critical factor in the prevention of spina bifida and other neural tube defects (Botto et al. 1999). Inadequate folate intake first leads to a decrease in serum folate concentration, then to a decrease in erythrocyte (red blood cell [RBC]) folate concentration, followed by a rise in homocysteine (Hcy) concentration and megaloblastic changes in bone marrow and other tissues with rapidly dividing cells (Institute of Medicine, Food and Nutrition Board 1999).

In our previous report (Colić Barić et al. in press), development of a folate-specific self-administered food frequency questionnaire (FFQ) was described and validated in

Correspondence: Irena Colić Barić, Faculty of Food Technology and Biotechnology, University of Zagreb, Pierottijeva 6, 10 000 Zagreb, Croatia. E-mail: icolic@pbf.hr

2 I. Colić Barić et al.

a population of adult women. The FFQ measures dietary folate equivalents (DFE) that express all forms of dietary folates, including synthetic folic acid used in fortified foods and dietary supplements, as an amount that is equivalent to naturally occurring food folate (Bailey 1998).

The aim of the present study was to assess the validity of the FFQ in a population with specific dietary patterns; namely, vegetarians. Vegetarians are expected to have superior folate status to omnivores due to higher vegetable consumption (Majchrzak et al. 2006). Folate is heat labile and has a short shelf-life, and fresh fruit and vegetables are excellent dietary sources (Kaić-Rak and Antičić 1990). Other important sources in a vegetarian diet include legumes and nuts.

Subjects and methods

Subjects

The subjects ($n = 75$) were vegetarians of both genders (13 males and 63 females) and varying dietary patterns (21 vegans and 54 lacto-ovo-vegetarians) (Table I). Veganism was defined as avoidance of any foods of animal origin, whereas lacto-ovo vegetarians consumed milk, eggs and dairy products (Position of the American Dietetic Association and Dietitians of Canada 2003).

Subjects were recruited through local vegetarian societies. Exclusion criteria included use of drugs known to interfere with folate metabolism and following a vegetarian diet for less than 12 months (Institute of Medicine, Food and Nutrition Board 1999). Participation was voluntary and all participants signed an informed consent form. The ethical committee of the Institute for Medical Research and Occupational Health approved the study protocol.

Food frequency questionnaire

The details of the FFQ were published previously (Colić Barić et al. in press). The FFQ is a 39-item questionnaire with the previous month as a reference period and the following available consumption frequencies: never, one/month, two to three/month, one/week, two to three/week, four to six/week and every day. The 39 separate food

Table I. Descriptive characteristics of the study population and biomarkers ($n = 75$).

Parameter	Mean	Standard deviation	Normal value
Age (years)	35.4	9.15	
Body mass index (kg/m ²)	22.0	3.09	
Females (% subjects)	82.7		
Males (% subjects)	17.3		
Vegans (% subjects)	28.0		
Lacto-ovo-vegetarians (% subjects)	72.0		
Current smokers (% subjects)	14.7		
Folate intake (µg DFE/day)	311.8	191.35	200 ^a
Serum folate (nmol/l)	28.8	6.19	16–35 ^b
Erythrocyte folate (nmol/l)	922.8	290.69	572–1843 ^b
Plasma Hcy (µmol/l)	9.8	2.75	5–15 ^b

^aRecommended folate intake for adults in Croatia (Ministarstvo zdravstva i socijalne skrbi 2004). ^bDefined by the manufacturer of the commercial kit (Abbott Laboratories, Diagnostics Division, Abbott Park, IL, USA).

65 items were dairy products (four items), eggs (one item), meat and fish (seven items),
legumes (three items), grain products (three items), vegetables (nine items), fruit
(seven items), cacao products (two items), vitamin drink (one brand most commonly
consumed in Croatia) and breakfast cereals (cornflakes and muesli). The subjects
70 received the FFQ in the form of a booklet with incorporated food photographs (Senta
et al. 2004). Folate intake was calculated using national food composition tables
taking into account retention factors since folate is heat labile (Kaić-Rak and AntoniĆ
1990; Agte et al. 2002). Beside food folate, folic acid originating from fortified foods
and dietary supplements was also assessed. At the moment, folate enrichment of, for
example, cereals is not mandatory and not practiced by Croatian food producers.

75 Subjects provided detailed information on supplement use and consumption of
folate-fortified food products currently available at the Croatian market; namely,
breakfast cereals, milk (two brands) and vitamin drinks. Self-reported dietary
supplement use (including folate) is considered valid and has been confirmed by
biological markers (Brantsaeter et al. 2007).

80 The FFQ measures DFE. The use of DFE is recommended for evaluating the
adequacy of folate intake (Suitor and Bailey 2000). To calculate the DFE, the folate
content of foods fortified with folate and the folate content of dietary supplements
were multiplied by 1.7 and the folate content of dietary supplements taken on an
empty stomach was multiplied by 2.

Biomarkers

85 Blood samples were drawn following an overnight fast in order to measure serum and
RBC folate and plasma Hcy. Folate and Hcy were determined using Abbott AxSYM
systems (Abbott Laboratories, Diagnostics Division, Abbott Park, IL, USA),
according to the manufacturer's instructions (Shipchandler and Moore 1995;
Manzella et al. 1999). Cut-off values for the folate level (16 and 572 nmol/l for
90 serum and RBC folate, respectively) were defined by the manufacturer of the
commercial kit and the cut-off for plasma Hcy was defined as 15 $\mu\text{mol/l}$ (Refsum et al.
2004). The values for serum and RBC folate represent a minimal normal value, and
the value for plasma Hcy represents a maximal normal value.

Statistical analyses

95 Several methods were used to assess the validity of the FFQ. A principal component
factor analysis was performed on three biomarkers. The variance of each variable in a
certain set of variables (in our study set of folate biomarkers) can be divided on
common variance (variance shared with other variables within the data-set) and
unique variance (the sum of specific variance and error variance). The first principal
100 component was calculated in order to represent as much as possible common variance
of a certain data-set. We assume that the common variance of folate biomarkers
represented by the first principal component is the best way to express overall folate
status. Pearson product-moment correlation coefficients were calculated for folate
intake based on the FFQ's and respective biomarkers of folate status. Sensitivity (the
105 ability of the FFQ to identify individuals with low folate status) and specificity (the
ability of the FFQ to identify individuals with adequate folate status) were calculated
using a 200 $\mu\text{g/day}$ cut-off value for folate intake, 15 $\mu\text{mol/l}$ for plasma Hcy, 16 nmol/l
for serum folate and 572 nmol/l for RBC folate. The ability of the FFQ to correctly

4 I. Colić Barić et al.

Table II. Principal components factor analysis of folate status biomarkers ($n=75$).

Factor structure matrix	Folate status factor
Serum folate	0.85
Erythrocyte folate	0.85
Plasma Hcy	-0.64
Eigenvalue	1.83
Total variance explained (%)	61

classify participants into quartiles of every respective biochemical indicator was also determined. Percentages of those correctly classified (participants classified within the same quartiles), closely classified (participants classified within the same or adjacent quartiles) and grossly misclassified (participants classified in the opposite quartiles) and the corresponding quadratic weighted kappa values were calculated.

STATISTICA 7.1 was used for the statistical analysis (StatSoft, Inc. 2006). The Kolmogorov–Smirnov test was used for testing the normality of data distribution. $P < 0.05$ was considered statistically significant.

Results

Plasma Hcy, serum, and RBC folate levels were normally distributed. The distribution of folate intakes estimated by the FFQ was right-skewed and was therefore log-transformed. The first principal component was highly correlated with all biomarkers (Table II). The Pearson product-moment correlation coefficients for biomarkers and folate intake were statistically significant for serum and RBC folate, but not for plasma Hcy (Table III).

Sensitivity and specificity of the folate FFQ for the cut-off value of 200 μg DFE are presented in Table IV. The 200 μg folate cut-off value was set as the recommended folate intake for adults in Croatia (Ministarstvo zdravstva i socijalne skrbi 2004). The quadratic weighted kappa for all biomarkers and folate status factor was above 0.2 and the gross misclassification of subjects into quartiles was less than 10% (Table V).

Discussion

Few FFQs have been created or modified to measure dietary intake among vegetarians (Waldmann et al. 2006) and, to our knowledge, this is the first study reporting the validity of a folate FFQ in vegetarians. The FFQ was already validated among adult women, but the FFQ is population specific and it is important to check its validity in

Table III. Pearson product-moment correlation coefficients with 95% confidence interval ($n=75$).

Parameter	Folate intake ($\log_{10}[\text{DFE } (\mu\text{g/day})]$)
Serum folate	0.41* (0.20–0.58)
Erythrocyte folate	0.36* (0.15–0.54)
Plasma Hcy	-0.15 (-0.36–0.08)
Folate status factor	0.41* (0.20–0.58)

* $P < 0.05$.

Validation of the folate FFQ in vegetarians 5

Table IV. Sensitivity and specificity (%) of the folate FFQ for cut-off value 200 μg DFE and absolute frequencies for each category ($n = 75$).

Parameter	Cut-off value	Folate intake		Sensitivity	Specificity
		<200 μg DFE	>200 μg DFE		
Serum folate	<16 nmol/l	0	1	0	69
	>16 nmol/l	23	51		
Erythrocyte folate	<572 nmol/l	3	5	38	70
	>572 nmol/l	20	47		
Plasma Hcy	>15 $\mu\text{mol/l}$	0	4	0	68
	<15 $\mu\text{mol/l}$	23	48		

vegetarians, who are usually excluded from the FFQ validation studies because of having a specific diet (Cade et al. 2002; Wise and Birrell 2002).

The use of biomarkers to validate dietary intake methods is increasing (Johnson et al. 2008). Validation was performed through comparison of folate intake and relevant biomarkers: serum and RBC folate and plasma Hcy. An advantage of using biomarkers is that they provide an objective measure of nutrient intake, with measurement errors essentially independent from those associated with dietary intake measures based on self report (van't Veer et al. 1993). Without the existence of the relevant biomarkers, the alternative would be relative validity; that is, comparison of the FFQ with other dietary assessment methods, which is the case with nutrients such as calcium (Subar 2004; Šatalić et al. 2007).

The RBC folate level is a good biomarker for folate status because of its correlation with the liver, a major folate storage site (Wu et al. 1975), and it reflects long-term intake (>3 months) (Jacob et al. 1998), whereas plasma and serum folate values are commonly accepted to reflect recent dietary intake. RBC folate is an indicator of long-term status because folate is absorbed only by the developing RBC in the bone marrow and not by the circulating mature RBC during its 120-day lifespan. The plasma concentration of Hcy (Krumdieck 1990) serves as an additional good indicator of folate nutritional status. Hcy increases when there is a deficiency of 5-methyl-tetrahydrofolate necessary to convert Hcy to methionine.

In this study, the correlation for folate intake based on the FFQ and serum and RBC folate did not differ significantly, although the correlation was higher for serum folate. This can be explained through the defined reference period of the FFQ, which was the previous month. Although it is not the most recent period, it is shorter than 3 months, which is a mature RBC's lifespan.

Table V. Cross-classification (%) into quartiles for folate intake and biomarkers and weighted kappa values ($n = 75$).

Folate intake	Same quartile	Same or adjacent quartile	Opposite quartile	Quadratic weighted kappa
Serum folate	36	77	4	0.39
Erythrocyte folate	33	77	4	0.37
Plasma Hcy	28	75	8	0.24
Folate status factor	41	83	7	0.42

160 The correlation of folate intake and Hcy level was negative as expected, but lower
than the correlation with serum and RBC folate, and not significant. When the FFQ
was administered for validation purposes among adult women (Colić Barić et al. in
165 **AQ1** press), the negative correlation between folate intake and plasma Hcy was significant.
The missing correlation in vegetarians can be explained by the high probability of their
inadequate B₁₂ status, which can elevate their Hcy in spite of adequate folate intake
(Stabler and Allen 2004). Also, it is generally agreed that plasma Hcy is most useful as
a functional measure of folate status when the concentration of folate in the diet is in
the low to moderate range, which is not the case in the vegetarian diet (O'Leary and
Sheehy 2002; Li et al. 2000).

170 When compared with recently published validation studies of folate FFQs, our
results compare well. One of the highest reported correlations of RBC folate with
folate intake (DFE) was 0.55 among pregnant Danish women (Mikkelsen et al. 2006).
A folate FFQ developed for the Dutch elderly showed a weak correlation between
folate intake and biomarkers ($r=0.14$, 0.05 and 0.02 for serum and RBC folate and
175 Hcy) but the FFQ was able to correctly classify subjects according to their folate
intake (van de Rest et al. 2007). Owens et al. (2007) reported a correlation for RBC
folate and folate intake (DFE) under 0.35 for the two questionnaires tested (DFE
screener and FFQ). The DFE screener sensitivity was 12% with a specificity of 91%.
The FFQ sensitivity was 12% with a specificity of 96%. Among Dutch women of
reproductive age, the FFQ was validated against serum and RBC folate (Verkleij-
180 Hagoort et al. 2007). Pearson correlations were 0.20 (FFQ versus serum folate) and
0.28 (FFQ versus RBC folate). Among adolescent females aged 16–19 years, the FFQ
was validated against serum and RBC folate (Green et al. 1998). For serum folate, the
FFQ classified 45% of the subjects correctly into quartiles, 80% were correctly and
closely classified, and 20% were misclassified (not within the same or adjacent
185 quartile). For RBC folate, the FFQ classified 32% of subjects correctly and 75%
correctly and closely, while 25% of subjects were misclassified. Among women
between the ages of 21 and 47 years, the correlation for folate FFQ and Hcy was –
0.26 (Yen et al. 2003).

190 Pearson's correlation, weighted kappa and classification into quartiles were more
favourable in vegetarians than among omnivore women in the previous validation
study (Colić Barić et al. in press). This is not surprising since vegetarians are more
health conscious and therefore more focused on personal diet, and it was shown that
195 vegetarians are able to recall their diet with higher reliability (Willett 1999; Kuzma and
Lindsted 1990).
AQ1

Based on the results of this validation study, it can be concluded that the folate FFQ
is a valid tool for measuring DFE intake in adult Croatian vegetarians.

Acknowledgements

The present study is a part of Scientific Project 058-0222411-2820, 'Diet, Homo-
cysteine and Bone Health', funded by The Ministry of Science, Education and Sport
200 of the Republic of Croatia.

Note

1 The FFQ may be obtained from the authors upon request.

References

- Agte V, Tarwadi K, Mengale S, Hinge A, Chiplonkar S. 2002. Vitamin profile of cooked foods: How healthy is the practice of ready-to-eat foods? *Int J Food Sci Nutr* 53:197–208.
- Bailey LB. 1998. Dietary reference intakes for folate: The debut of dietary folate equivalents. *Nutr Rev* 56:294–249.
- Bailey LB, Gregory JF. 2006. Folate. In: Bowman BA, Russell RM, editors. *Present knowledge in nutrition*. 9th ed. Washington, DC: ILSI. pp 278–302.
- Botto LD, Moore CA, Khoury MJ, Erickson JD. 1999. Neural-tube defects. *N Engl J Med* 341:1509–1519.
- Brantsaeter AL, Haugen M, Hagve TA, Aksnes L, Rasmussen SE, Julshamn K, Alexander J, Meltzer HM. 2007. Self-reported dietary supplement use is confirmed by biological markers in the Norwegian Mother and Child Cohort Study (MoBa). *Ann Nutr Metab* 51:146–154.
- Cade J, Thompson R, Burley V, Warm D. 2002. Development, validation and utilisation of food-frequency questionnaires—a review. *Public Health Nutr* 5:567–587.
- AQ1** Colić Barić I, Šatalić Z, Keser I, Ceci I, Sučić M. In press. Validation of the folate food frequency questionnaire with serum and erythrocyte folate and plasma homocysteine. *Int J Food Sci Nutr*.
- Green TJ, Allen OB, Oconnor DL. 1998. A three-day weighed food record and a semiquantitative food-frequency questionnaire are valid measures for assessing the folate and vitamin b-12 intakes of women aged 16 to 19 years. *J Nutr* 128:1665–1671.
- Institute of Medicine, Food and Nutrition Board. 1999. *Dietary reference intakes for thiamin, riboflavin, niacin, vitamin B6, folate, vitamin B12, pantothenic acid, biotin, and choline*. Washington, DC: National Academy Press.
- Jacob RA, Gretz DM, Taylor PC, James SJ, Pogribny IP, Miller BJ, Henning SM, Swendseid ME. 1998. Moderate folate depletion increases plasma homocysteine and decreases lymphocyte DNA methylation in postmenopausal women. *J Nutr* 128:1204–1212.
- AQ2** Johnson RK, Yon BA, Hankin JH. 2008. Dietary Assessment and validation. In: Mosen ER, Van Horn L, editors. *Research: successful approaches*. 3rd ed. American Dietetic Association. pp 187–204.
- Kaić-Rak A, Antonić K. 1990. *Tablice o sastavu namirnica i pića*. Zagreb: Zavod za zaštitu zdravlja SR Hrvatske.
- Krumdieck CL. 1990. Folic acid. In: Brown ML. editor. *Present knowledge in nutrition*. 6th ed. Washington, DC: ILSI. pp 179–188.
- Kuzma JW, Lindsted KD. 1990. Determinants of eight-year diet recall ability. *Epidemiology* 1:386–391.
- Li D, Sinclair AJ, Mann NJ, Turner A, Ball MJ. 2000. Selected micronutrient intake and status in men with differing meat intakes, vegetarians and vegans. *Asia Pac J Clin Nutr* 9:18–23.
- Life Sciences Research Office. 1989. *Nutrition monitoring in the United States: An update report on nutrition monitoring*. Federation of American Societies for Experimental Biology. Washington, DC: US Department of Health and Human Services, Public Health Service.
- Majchrzak D, Singer I, Maenner M, Rust P, Genser D, Wagner KH, Elmadsfa I. 2006. B-vitamin status and concentrations of homocysteine in Austrian omnivores, vegetarians and vegans. *Ann Nutr Metab* 50:485–491.
- Manzella S, Gronowski A, Ladenson J, Scott MG. 1999. Limited linear range of the Abbott AxSYM serum and erythrocyte folate methods. *Clin Chem* 45:582–583.
- Mikkelsen TB, Osler M, Olsen SF. 2006. Validity of protein, retinol, folic acid and n-3 fatty acid intakes estimated from the food-frequency questionnaire used in the Danish National Birth Cohort. *Public Health Nutr* 9:771–778.
- Ministarstvo zdravstva i socijalne skrbi [Ministry of Health and Social Welfare]. 2004. *Pravilnik o hrani za posebne prehrabene potrebe [Regulation of food for specific dietary purposes]*. Narodne Novine 81. Zagreb: Narodne Novine Corporation.
- O’Leary K, Sheehy PJA. 2002. Plasma, liver and kidney folate and plasma homocysteine concentrations are poor response variables at very low dietary folate intakes, in a folate depletion/repletion rat model. *Int J Food Sci Nutr* 53:35–42.
- Owens JE, Holstege DM, Clifford AJ. 2007. Comparison of two dietary folate intake instruments and their validation by RBC folate. *J Agric Food Chem* 55:3737–3740.
- Position of the American Dietetic Association and Dietitians of Canada. 2003. Vegetarian diets. *J Am Diet Assoc* 103:748–765.
- Refsum H, Smith AD, Ueland PM, Nexø E, Clarke R, McPartlin J, Johnston C, Engbaek F, Schneede J, McPartlin C, Scott JM. 2004. Facts and recommendations about total homocysteine determinations: An expert opinion. *Clin Chem* 50:3–32.

8 I. Colić Barić et al.

- Šatalić Z, Colić Barić I, Cecić I, Keser I. 2007. Short food frequency questionnaire can discriminate inadequate and adequate calcium intake in Croatian postmenopausal women. *Nutr Res* 27:542–547.
- Senta A, Pucarín-Cvetković J, Doko Jelinić J. 2004. Kvantitativni modeli namirnica i obroka. Zagreb: Medicinska naklada.
- Shipchandler MT, Moore EG. 1995. Rapid, fully automated measurement of plasma homocysteine with the Abbott IMx analyzer. *Clin Chem* 41:991–994.
- Stabler SP, Allen RH. 2004. Vitamin B12 deficiency as a worldwide problem. *Annu Rev Nutr* 24:299–326.
- StatSoft, Inc. 2006. STATISTICA data analysis software system. version 7.1. Available online at: www.statsoft.com.
- Subar AF. 2004. Developing dietary assessment tools. *J Am Diet Assoc* 104:769–770.
- Suitor CW, Bailey LB. 2000. Dietary folate equivalents: Interpretation and application *J Am Diet Assoc* 100:88–94.
- van de Rest O, Durga J, Verhoef P, Melse-Boonstra A, Brants HAM. 2007. Validation of a food frequency questionnaire to assess folate intake of Dutch elderly people. *Br J Nutr* 98:1014–1020.
- van't Veer P, Kardinaal AF, Bausch-Goldbohm RA, Kok FJ. 1993. Biomarkers for validation. *Eur J Clin Nutr* 47(Suppl 2):58–63.
- Verkleij-Hagoort AC, de Vries JHM, Stegers MPG, Lindemans J, Ursem NTC, Steegers-Theunissen RPM. 2007. Validation of the assessment of folate and vitamin B12 intake in women of reproductive age: the method of triads. *Eur J Clin Nutr* 61:610–615.
- Wagner C. 1996. Symposium on the subcellular compartmentation of folate metabolism. *J Nutr* 126:1228S–1231S.
- Waldmann A, Dorr B, Koschizke JW, Leitzmann C, Hahn A. 2006. Dietary intake of vitamin B-6 and concentration of vitamin B-6 in blood samples of German vegans. *Public Health Nutr* 9:779–784.
- Willett WC. 1999. Convergence of philosophy and science: The Third International Congress on Vegetarian Nutrition. *Am J Clin Nutr* 70:434S–438S.
- Wise A, Birrell NM. 2002. Design and analysis of food frequency questionnaires—review and novel method. *Int J Food Sci Nutr* 53:273–279.
- Wu A, Chanarin I, Slavin G, Levi AJ. 1975. Folate deficiency in the alcoholic—its relationship to clinical and haematological abnormalities, liver disease and folate stores. *Br J Haematol* 29:469–478.
- Yen J, Zoumas-Morse C, Pakiz B, Rock CL. 2003. Folate intake assessment: Validation of a new approach. *J Am Diet Assoc* 103:991–1000.

AQ3